

WEICKMANN & WEICKMANN, Postfach 860 820, 81635 München

European Patent Office

80298 Munich

DIPL.-ING. H. WEICKMANN (bis 31.1.01)
 DIPL.-ING. F. A. WEICKMANN
 DIPL.-CHEM. B. HUBER
 DR.-ING. H. LISKÄ
 DIPL.-PHYS. DR. J. PRECHTEL
 DIPL.-CHEM. DR. B. BOHM
 DIPL.-CHEM. DR. W. WEISS
 DIPL.-PHYS. DR. J. TIESMEYER
 DIPL.-PHYS. DR. M. HERZOG
 DIPL.-PHYS. B. RUTTENSBERGER
 DIPL.-PHYS. DR.-ING. V. JORDAN
 DIPL.-CHEM. DR. M. DEY
 DIPL.-FORSTW. DR. J. LACHNIT

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DATUM / DATE

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Application No./Patent No.:

PCT/EP 03/04650

Applicant/Proprietor:

DeveloGen Aktiengesellschaft für entwicklungsbiologische Forschung

Encl.

Sequence Listing (diskette) ✓

Sequence Listing (paper
copy) (all 1-fold) ✓New description pages 60 ✓
and 61 ✓

In response to the official communication
dated June 26, 2003.

Applicants herewith submit a sequence listing
in computer-readable form as well as in the
form of a paper copy.

It is stated that the information recorded on
the data carrier is identical to the written
sequence listing. It is further stated that the
sequence listing does not include subject
matter which goes beyond the content of the
application as originally filed.

Postfach 860 820
81635 München

Kopernikusstraße 9
81679 München

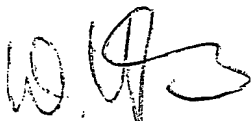
Deutschland

Telefon (089) 45563 0
 (0700) WEICKMAN
 Telefax (089) 45563 999
 E-Mail email@weickmann.de
 Internet www.weickmann.de
 Vat-ID. Nr. DE 130 753 315

Hypovereinsbank München
 Konto 208 401 (BLZ 700 202 70)
 S.W.I.F.T.-Adresse HYVE DE MM

Postbank München
 Konto 77 46-804 (BLZ 700 100 80)
 S.W.I.F.T.-Adresse PBNKDEFF 700

Furthermore, new description pages 60 and 61 are filed including the correct sequence numberings.



Dr. W. Weiss

temperature (preferably 22°C), 40 per cent humidity and a light / dark cycle of preferably 14 / 10 hours. The mice were fed a standard chow (for example, from ssniff Spezialitäten GmbH, order number ssniff M-Z V1126-000). For the fasting experiment ("fasted wild type mice"), wild type mice were starved for 48 h without food, but only water supplied ad libitum (see, for example, Schnetzler et al., (1993) J Clin Invest 92(1):272-280, Mizuno et al., (1996) Proc Natl Acad Sci U S A 93(8):3434-3438). Animals were sacrificed at an age of 6 to 8 weeks. The animal tissues were isolated according to standard procedures known to those skilled in the art, snap frozen in liquid nitrogen and stored at -80°C until needed.

RNA was isolated from mouse tissues using Trizol Reagent (for example, from Invitrogen, Karlsruhe, Germany) and further purified with the RNeasy Kit (for example, from Qiagen, Germany) in combination with an DNase-treatment according to the instructions of the manufacturers and as known to those skilled in the art. Total RNA was reverse transcribed (preferably using Superscript II RNaseH- Reverse Transcriptase, from Invitrogen, Karlsruhe, Germany) and subjected to Taqman analysis preferably using the Taqman 2xPCR Master Mix (from Applied Biosystems, Weiterstadt, Germany; the Mix contains according to the Manufacturer for example AmpliTaq Gold DNA Polymerase, AmpErase UNG, dNTPs with dUTP, passive reference Rox and optimized buffer components) on a GeneAmp 5700 Sequence Detection System (from Applied Biosystems, Weiterstadt, Germany).

Taqman analysis was performed preferably using the following primer/probe pairs:

For the amplification of Sac domain-containing inositol phosphatase 2 (sac2) (SEQ ID NO:2): 5'- CCT GGA TCG CAC CAA CG -3'; mouse sac2 reverse primer (SEQ ID NO:3): 5'- TTA AGC TGC TGT TCC ATG ACC A

-3'; Taqman probe (SEQ ID NO:4): (5/6-FAM) TCC AGG CTG CCA TAG
CGC GC (5/6-TAMRA)

For the amplification of mouse solute carrier family 25 (mitochondrial
5 carrier, Aralar) member 12 (Slc25a12) (SEQ ID NO:5): 5'- CCT GCC AAC
CCT GAT CAC A -3'; mouse Slc25a12 reverse primer (SEQ ID NO:6): 5'-
TTT CAA TGC CAG CGA AAG TG -3'; Taqman probe (SEQ ID NO:7):
(5/6-FAM) CGG TGG CTA CAG ACT TGC CAC GG (5/6-TAMRA)

10 For the amplification of mouse solute carrier family 25 (mitochondrial
carrier; adenine nucleotide translocator), member 13 (Slc25a13) (SEQ ID
NO: 8): 5'- AGC GGT GGT TCT ATG TCG ATT T -3'; mouse Slc25a13
reverse primer (SEQ ID NO: 9): 5'- CGG GAT TTA GGA ACC GGC T -3';
Taqman probe (SEQ ID NO:10): (5/6-FAM) AGG CGT GAA GCC CGT GGG
15 ATC T (5/6-TAMRA)

For the amplification of mouse myelin gene expression factor 2 (mef2)
(SEQ ID NO: 11): 5'- ACA AGG ATG GCA AGA GCA GAG -3'; mouse
mef2 reverse primer (SEQ ID NO: 12): 5'- ATG GAA ATT GCT TGG ACT
20 GCT T -3'; Taqman probe (SEQ ID NO: 13): (5/6-FAM) CAT GGG CAC
TGT CAC TTT TGA GCA GG (5/6-TAMRA)

In the figures the relative RNA-expression is shown on the Y-axis. In
Figures 4A and B, 8A, B, C, and D, and 16A, B, and C, the tissues tested
25 are given on the X-axis. "WAT" refers to white adipose tissue, "BAT"
refers to brown adipose tissue.

As shown in Figure 4A, real time PCR (Taqman) analysis of the expression
of the Sac domain-containing inositol phosphatase 2 (SAC2) RNA in
30 mammalian (mouse) tissues revealed that SAC2 is highly expressed in
hypothalamus, brain, WAT, spleen and kidney. Figure 4B shows that SAC2
is upregulated in BAT and pancreas of fasted animals as well as ob / ob